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Flavonol glycosides and seed coat structure in certain species of *Epilobium* – A correlation?¹

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Summary. In a survey of 239 populations of *Epilobium* representing 17 taxa the following flavonol glycosides were found: myricetin 3-O-arabinoside; 3-O-glucoside; 3-O-rhamnoside; quercetin 3-O-arabinoside; 3-O-glucoside; 3-O-diglucoside; 3-O-rhamnoside; kaempferol 3-O-glucoside; and 3-O-rhamnoside. A correlation appears to exist between seed coat sculpturing as determined in a previous study using SEM techniques, and the flavonoid profiles.

The genus *Epilobium* is quite cosmopolitan in distribution, with several circumboreal species². In many respects it appears to be ecologically nonspecific, but at the same time exhibiting a wide range of morphological and cytological variation. This is shown to its greatest degree in western North America³. The present day distribution of the genus indicating a possible centre of origin in this region⁴.

With this in mind a survey of 17 taxa representative of the genus in northwestern America was carried out to determine if a parallel complexity occurred with respect to leaf flavonoid glycosides.

In the Onagraceae, in general, studies have been restricted mainly to the genera *Oenothera*⁵, *Circaea*⁶, with little being carried out on other taxa. Studies have shown that the commonly distributed flavonol glycosides⁵ are present, and

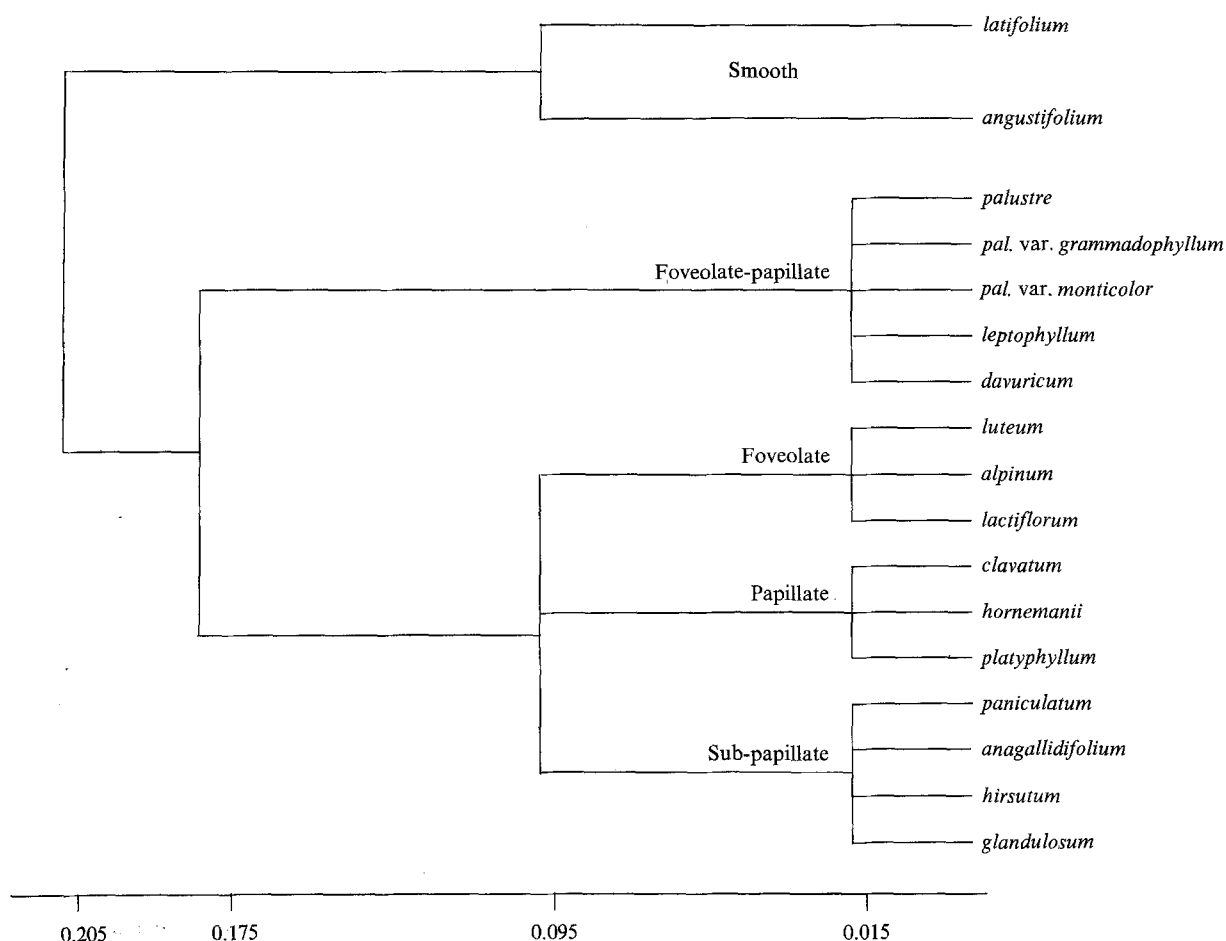
rarely more complicated flavonoids such as methylated derivatives and glyco-flavones⁶.

In the present study the leaves of 239 populations were sampled representing 17 taxa (table) previously studied for their seed coat structures⁷. A minimum of 6 populations (10 g dry wt) were examined in any taxon. Isolation of flavonoid glycosides, hydrolytic procedures, and spectral analyses were carried out using standard methods⁸, with modifications when discontinuities in distribution of compounds was found. In such situations flavonoids were exhaustively extracted using 70% methanol reduced to dryness and loaded in a minimum volume of water onto a sephadex LH 20 column. Gradient elution of glycoside mixtures was then carried out using water-methanol-acetone gradients. Flavonoids were then re-isolated and characterized using normal analytical procedures.

Flavonol glycosides in *Epilobium*

Taxon	Samples	m 3 glc	m 3 rh	m 3 arab	q 3 glc	q 3 rh	q 3 arab	q 3 diglc	k 3 glc	k 3 rh
<i>E. latifolium</i>	66	+	+	+	+	+	+	+	+	+
<i>E. angustifolium</i>	52	+	+	+	+	+	+		+	48
<i>E. glandulosum</i>	6	+	+	+	+	+	+			+
<i>E. paniculatum</i>	8	+	+	6	+	+	7			7
<i>E. hirsutum</i>	8	+	+	+	+	+	+			7
<i>E. anagallidifolium</i>	8	+	+	7	+	+	+			+
<i>E. platyphyllum</i>	6	+	+		+	+	+			+
<i>E. hornemanii</i>	12	+	+		+	+	10			+
<i>E. clavatum</i>	6	+	5		+	+	5			+
<i>E. lactiflorum</i>	6	+	+		+	+	+			
<i>E. alpinum</i>	17	+	+		+	+	+			
<i>E. luteum</i>	6	+	+		+	+	4			
<i>E. davuricum</i>	6	+			+	+				
<i>E. leptophyllum</i>	6	+			+	+				
<i>E. palustre</i>	14	+			+	+				
<i>E. palustre</i> var. <i>grammadophyllum</i>	12	+			+	+				
<i>E. palustre</i> var. <i>monticolor</i>	10	+			+	+				
Total	239									

m, myricetin; glc, glucose; q, quercetin; arab, arabinose; k, kaempferol; rh, rhamnose.



Epilobium single linkage analysis of flavonoid glycosides.

9 flavonol glycosides were isolated (table) based on myricetin, quercetin and kaempferol and the sugars glucose, arabinose, and rhamnose. All were simple 3-O-glycosides, however, their distributions were not ubiquitous within each taxon investigated and certain glycosides showed both inter and intra specific variation. An example of this occurs in *E. paniculatum* and *E. anagallidifolium* in which myricetin 3-O-arabinoside occurs in 75 and 85% of the populations respectively. From a phylogenetic viewpoint a significant observation in the absence of kaempferol glycosides from 8 of the taxa investigated (table). Harborne⁹ had indicated that an evolutionary relationship exists which parallels reduction in 6-ring hydroxylation (myricetin → quercetin → kaempferol; primitive → advanced). Such an observation in this group may be worth pursuing from this aspect.

Previous studies by the present author using seed coat structure⁷ (SEM) indicated a subdivision of the genus into papillate → foveolate → to nonpapillate groupings. A single linkage analysis of the flavonoid profiles (figure) indicates that this subdivision holds true with respect to flavonoids also. 1 group (*E. palustre*, etc.) was previously characterized as subpapillate (does not possess myricetin 3-O-rhamnoside), however, a single linkage analysis separates this group from the *E. paniculatum* group and it allies, a subgroup which was also designated subpapillate and has a chemical profile which includes myricetin 3-O-rhamnoside. On re-examining the papillation present it was concluded

that 2 types of subpapillate forms exist, the *E. paniculatum* type being best described as foveolate-papillate.

In conclusion, it appears that a positive correlation exists between the leaf of flavonoid profiles of the taxa investigated and their seed coat structures⁷. However, despite the morphological and cytological variation found in these species in northwestern North America, very little variation, and a marked lack of flavonoid complexity, paradoxically typifies the genus.

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